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The metabolic fate of alpha linolenic acid (ALA)

Extremely limited conversion efficiency

By Leah Gillingham, MSc, PhD

ABSTRACT:

EPA and DHA possess important physiological and biological properties in human health and development; however, it is alpha-linolenic acid (ALA) that is classified as the essential n-3 PUFA. While humans have the required enzymes to biosynthesize omega-3 long-chain polyunsaturated fatty acids (PUFA), studies demonstrate that the majority of ALA is β -oxidized and only ~5% of ALA is converted to eicosapentaenoic acid (EPA) and <0.5% of ALA is converted to docosahexaenoic acid (DHA). Even very high intakes of dietary ALA fail to effectively modulate plasma and tissue levels of DHA. Furthermore, the abundance of linoleic acid (LA) in our diet, as well as age, gender and genetics influence the conversion efficiency of ALA. Therefore, direct dietary consumption or supplementation of EPA and DHA from fish, fish oil, or algae is the only clinically effective way to increase blood and tissue levels of these long-chain PUFA in humans for optimal health and disease prevention.

INTRODUCTION

Considerable clinical interest has focused on the health benefits of omega-3 polyunsaturated fatty acids (ω -3 PUFA). While consumption of marine derived eicosapentaenoic acid (EPA; 20:5 ω -3) and docosahexaenoic acid (DHA; 22:6 ω -3) have been evidenced to prevent cardiovascular and neurological disease risk, the specific function of plant derived alpha-linolenic acid (ALA; 18:3 ω -3) remains a matter of debate. EPA and DHA play a vital role in cellular membranes, maintaining fluidity, protein and cellular functions, as well as influencing eicosanoid metabolism, gene expression and cell signaling (Adkins 2010). However, the specific bioactive role of ALA is unclear (Sinclair 2002). It has been suggested that the major function of ALA is to serve as a precursor for EPA and DHA, a pathway that has received much attention and clinical investigation.

In 1929, Burr and Burr first identified the nutritional essentiality of LA, and later ALA (Burr 1929, Burr 1930). ALA is the parent ω -3 PUFA containing three double bonds with the first double bond located at

the third carbon relative to the methyl end of the 18-carbon chain. ALA and ω -6 linoleic acid (LA, 18:2 ω -6) are termed essential fatty acids because humans lack the delta (Δ)15- and Δ 12-desaturase enzymes required for insertion of a double bond at the ω -3 or ω -6 position, respectively. Since EPA and DHA can be synthesized from ALA via a series of alternating desaturation and elongation steps, these long-chain PUFA are not considered essential. However, whether endogenous synthesis of EPA and DHA from ALA is adequate to support growth, physiological needs, and disease risk reduction is questioned (Harris 2009, Saldanha 2009). Considering negligible plasma and tissue levels of ALA, yet its classification of essential, this review will examine the biosynthesis of EPA and DHA from ALA, factors influencing ALA metabolism, and ω -3 PUFA in the current diet.

BIOSYNTHESIS OF LONG-CHAIN POLYUNSATURATED FATTY ACIDS

The predominate site of ALA desaturation and elongation occurs in the liver, however, also occurs to a lesser extent in other tissues, including the brain and heart (Cho 1999 A, Cho 1999 B). Dietary LA and ALA compete for the same desaturase and elongase enzymes for long-chain PUFA biosynthesis with the majority of this pathway occurring in the endoplasmic reticulum. Of interest, the desaturase enzymes have a higher affinity for ALA versus LA (Plourde 2007), however, high levels of dietary LA saturate Δ 6-desaturation inhibiting the accumulation of ω -3 long-chain PUFA, namely EPA (Angela Liou 2009, Liou 2007).

The first reaction in the conversion pathway is the desaturation of ALA to stearidonic acid (SDA; 18:4 ω -3) or LA to gamma-linolenic acid (GLA; 18:3 ω -6) via the rate-limiting enzyme Δ 6-desaturase (Sprecher 2000) (Figure 1). Next, elongation and Δ 5-desaturation converts SDA to EPA and GLA to arachidonic acid (AA; 20:4 ω -6), the major bioactive n-6 PUFA in tissue membranes and precursor for proinflammatory eicosanoids (Adkins 2010). Compared to AA, EPA is quantitatively a minor fatty acid in tissue membranes and undergoes further elongation to docosapentaenoic acid (DPA; 22:5 ω -3), or is metabolized by cyclooxygenase (COX) and lipoxygenase (LOX) enzymes in the synthesis of anti-inflammatory eicosanoids. In humans, DPA undergoes another elongation and Δ 6-desaturation step, and then partial β -oxidation in the peroxisome to form DHA (Sprecher 2000). Structurally, **DHA is the predominant ω -3 PUFA esterified into tissue membrane phospholipids.** DHA can also be

metabolized to produce resolvins and protectins in the resolution of inflammation (Adkins 2010). It has been hypothesized that multiple use of the rate-limiting $\Delta 6$ -desaturase enzyme for the conversion of ALA to SDA and 24:5n-3 to 24:6n-3 may lead to a “bottle-neck” in the metabolic pathway and an associated decrease in the synthesis of DHA (D’andrea 2002, Kitson 2010). Another possible rate-limiting step may be related to the compartmental translocation of 24:6n-3 from the endoplasmic reticulum to the peroxisome. Both hypotheses demand further investigation.

RESULTS FROM HUMAN STUDIES:

The conversion of ALA to long-chain ω -3 PUFA is very inefficient. Dietary supplementation and isotope tracer studies in humans demonstrate a direct linear relationship between dietary ALA and plasma and tissue concentrations of EPA, at approximately 5% conversion efficiency (Plourde 2007). However, biosynthesis of DHA is negligible with typically less than 0.5% conversion of ALA to DHA. Human studies supplementing ALA ranging from 1.5 to 40 g/day for a minimum of three weeks report a relative increase in phospholipid concentration of EPA ranging from trace levels to 250% (Plourde 2007). Studies have also observed an increase in plasma and tissue concentrations of DPA, although to a lesser extent than EPA concentrations. However, the majority of dietary ALA intervention studies fail to effectively modulate plasma and tissue levels of DHA. Gillingham et al demonstrated that consuming 20 g/day of ALA from a flaxseed oil supplemented diet (~2.5 tbsp flaxseed oil daily) resulted in a relative 222% increase in plasma EPA concentration (1.74% total fatty acids) compared with control (0.54% total fatty acids) (2011). However, DHA concentration did not change after supplementing with flaxseed oil. Similarly, supplementing lactating women with 20 g/day of flaxseed oil (10.7 g/day ALA) for four weeks increased breast milk EPA and DPA, but not DHA concentrations (Francois 2003).

Additionally, very high levels of dietary ALA, as well as LA, may saturate the $\Delta 6$ -desaturase enzyme further inhibiting the final steps of $\Delta 6$ -desaturation of 24:5n-3 to DHA (Gibson 2011). Even with vegetarian and vegan diets, ALA conversion to DHA is not up-regulated (Fokkema 2000, Li 1999). James et al revealed that compared to dietary ALA, supplementing with SDA-enriched vegetable oil (therefore, bypassing the first $\Delta 6$ -desaturase rate-limiting step) more effectively increased plasma and tissue concentrations of EPA, however, did not raise DHA concentrations (2003). In addition, EPA supplementation does not significantly elevate DHA concentrations (Brenna 2009). Therefore, the only effective way to increase tissue DHA concentrations is through its direct consumption.

FACTORS AFFECTING EPA AND DHA BIOSYNTHESIS FROM ALA

DIETARY Ω -6 LINOLEIC ACID CONTENT:

The abundance of LA and resulting high ω -6 to ω -3 PUFA ratio in current Western diets significantly impedes the conversion efficiency of ALA. Liou and colleagues demonstrated that while maintaining dietary ALA at 1% energy, increasing levels of dietary LA from 4% to 10% energy, thus varying the ω -6 to ω -3 PUFA ratio from 4:1 to 10:1, resulted in a decrease in plasma EPA concentrations in healthy men (Angela Liou 2009, Liou 2007). In another human dietary intervention study, lowering the ω -6 to ω -3 PUFA ratio resulted in a significant decrease in platelet aggregation (Freese 1994). Furthermore, trans fats rich in hydrogenated vegetable oils inhibit the synthesis of ω -3 long-chain PUFA (Kummerow 2004).

MICRONUTRIENTS: Vitamins B3, B6, and C, magnesium and zinc are important cofactors to the $\Delta 5$ - and $\Delta 6$ -desaturase enzymes. Therefore, low intakes of these essential micronutrients may lead to a reduction in the synthesis of ω -3 long-chain PUFA (Cunnane 1988, de Lorgeril 2005, Harris 2009).

GENDER: Gender may affect biosynthesis of long-chain PUFA, as DHA concentration of plasma phospholipids have been shown to be higher in women than in men

(Decsi 2011). Giltay et al observed a 15% increase in DHA status in women compared with men (2004). Furthermore, administration of oral estradiol increased DHA status by 42%, while testosterone decreased DHA status by 22%. It is proposed that estrogen may upregulate ALA metabolism to DHA, and thus, increase maternal DHA status particularly during pregnancy due to the greater demand of DHA for fetal neurological development (Otto 2001, Innis 2000).

AGE: Studies suggest that infants exhibit increased conversion of ALA to long-chain PUFA, including DHA

(Clark 1992, Jensen 1996). A study tracing ALA metabolism in preterm infants fed long-chain PUFA enriched formula reported that in one-month old infants about 42% of ALA was converted to DHA, whereas only 11% was converted at three-months of age, and 7% at seven months of age (Carnielli 2007). Typically, conversion of ALA to DHA is still limited in healthy infants at ~1% (Brenna 2009). However, differences in adult age (18–29 versus 45–69 years) have not been shown to influence metabolism of ALA to EPA or DHA (Patenaude 2009).

GENETICS: Fatty acid desaturase genes FADS1 and FADS2 encode for $\Delta 5$ -desaturase and $\Delta 6$ -desaturase, respectively (Cho 1999, Cho 1999). Schaeffer and colleagues published the first study reporting that single nucleotide polymorphisms (SNP) of the FADS1/FADS2 gene cluster modulate ALA and LA metabolism

leading to differences in phospholipid PUFA concentrations (2006). More specifically, minor allele carriers for SNPs located in the FADS1/FADS2 gene cluster have reduced ability to convert LA to AA (Tanaka 2009) or ALA to EPA (Gillingham 2013a). Of interest, genetic variation explains ~40% or more of the interindividual variability in all fatty acid concentrations (Lemaitre 2008). Martinelli et al reported that individuals carrying FADS polymorphisms, associated with higher conversion of LA to AA, have increased proinflammatory CRP concentrations and risk for coronary artery disease (2008).

OTHER METABOLIC PATHWAYS

β -oxidation accounts for the major metabolic fate of dietary ALA (McCloy 2004). McCloy et al. demonstrated that ~71% of dietary ALA was oxidized over a seven day period, the highest oxidation rate of all fatty acids (2004). During mitochondrial β -oxidation of ALA, carbon units generated in the form of acetyl-CoA can be recycled to synthesize SFA, MUFA, cholesterol and ketone bodies (Burdge 2003). Second to β -oxidation, storage of ALA in adipose tissues accounts for a main metabolic route of dietary ALA (McCloy 2004), however, ALA is readily mobilized from fatty acid tissue during increased energy demands of the body.

DIETARY OMEGA-3 POLYUNSATURATED FAT INTAKE AND RECOMMENDATIONS

Since ALA and LA are the only fatty acids classified as essential, the US Institute of Medicine's (IOM) Food and Nutrition Board, together with Health Canada, has established an Adequate Intake (AI, an intake level necessary to achieve nutritional adequacy and prevent deficiency symptoms) for ALA as 1.1-1.6 g/day and for LA as 12-17 g/day for adults (Institute of Medicine 2002). Symptoms of ALA and LA deficiency include scaly dry skin, reproductive failure, numbness, weakness, pain, and blurred vision (Collins 1971, Holman 1982). However, current Western intake of ALA at 1.4-1.8 g/day and LA at 13-18 g/day by adults (Rhodes 2012) exceed the outlined AI by the IOM. The clinical concern is that current intake of combined EPA and DHA at 90-120 mg/day fail to meet recommendations outlined by professional health organization ranging from 500-4000 mg/day (Table 1). Due to the importance of DHA in fetal retina and brain growth and development, the minimum recommended intake of DHA during pregnancy and lactation is 200 mg/day (Koletzko 2007). However, pregnant and lactating women in Canada and the United States only consume

approximately 80 and 50 mg/day, respectively (Denomme 2005, Rhodes 2012). In addition, biosynthesis of long-chain PUFA, namely DHA from dietary ALA is insufficient to meet target recommended intakes of EPA and DHA for optimal health and disease prevention.

The FAO/WHO joint committee recommends an ω -6 to ω -3 ratio between 5:1 and 10:1 (WHO/FAO 1995). Of interest, the Paleolithic diet contained an ω -6 to ω -3 ratio of <1:1 (Kuipers 2010), while current intakes range from 10-25:1 in the North America diet.

SAFETY OF EPA AND DHA SUPPLEMENTATION

In 1997, the US Food and Drug Administration (FDA) granted Generally Recognized As Safe (GRAS) status to refined fish oils and indicated that the consumption of up to 3g/day of EPA+DHA is considered safe for the adults population, including patients with diabetes, bleeding tendencies, and elevated LDL-cholesterol (1997). Intake exceeding 3g/day should only be recommended under the guidance of a healthcare practitioner.

DIETARY SOURCES:

ω -6 LA is abundant in our diet, rich in corn, safflower, soybean, and sunflower oils. However, ω -3 ALA is found in only a limited amount of foods, namely flaxseed, walnuts, soybean and their oils, as well as canola oil, butternuts, and chia seeds. Although flaxseed oil represents the richest source of ALA (7.3 g ALA/tbsp), it is not commonly consumed compared with soybean oil (0.9 g ALA/tbsp) and canola oil (1.3 g ALA/tbsp) (USDA 2011). Purslane, a wild leafy vegetable common in the Eastern Mediterranean diet, contains 300-400mg ALA/100g serving (Simopoulos 1996).

EPA and DHA are primarily found in fish, fish oil, krill oil or algae. However, the concentration of EPA and DHA substantially varies in fish species (Table 2). For example, fatty fish such as farmed Atlantic salmon provides 1.83 g EPA+DHA per 3 oz serving, whereas the same amount of lean fish such as cod provides only 0.14 g EPA+DHA (USDA 2011). Therefore, one would have to consume approximately four servings of farmed Atlantic salmon or 52 servings of cod per week to achieve a recommended intake of 1g/day (Table 2). Where fatty fish intake exceeds two servings per day, such as Greenland and Japan (Stark 2002, Iso H 1989), plasma EPA/DHA levels are in what is widely considered a therapeutic range (Harris 2010). However, in most parts of the world, these

levels are not being achieved. In addition, depending on the species of fish, excessive fish intake may lead to concerns surrounding mercury exposure, particularly in pregnancy (Koletzko 2007). Taken together, fish oil supplements may provide a more feasible option to target clinical recommendations for EPA+DHA. Minor amounts of EPA can also be found in seaweed, such as kelp, laver, and wakame ranging in EPA levels from 0.004 to 0.186 g/100g serving (USDA 2011). Additionally, microalgae-based supplements offer an alternative for DHA intakes for vegetarians/vegans.

CONCLUSION

The consensus of human clinical trials substantiate that the biosynthesis of EPA and DHA from dietary ALA is extremely limited and insufficient to meet protective tissue levels for disease prevention. In addition, factors including diet, gender, age and genetics affect individual's capacity for biosynthesis and resulting EPA and DHA tissue concentration. Considering the majority of dietary ALA is β -oxidized, negligible conversion rates, and low intakes of EPA and DHA in the current diet, professional organizations emphasize direct supplementation of EPA and DHA in the diet for optimal health and disease risk reduction.

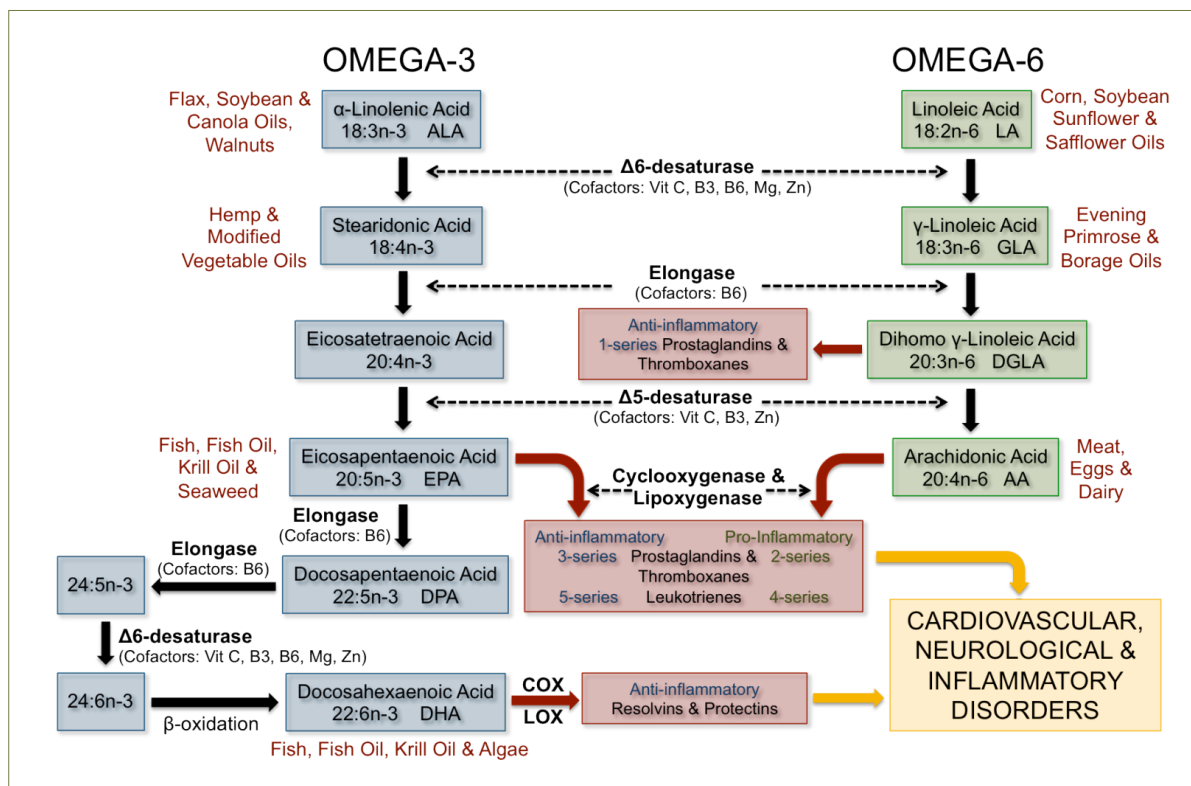


Figure 1: Metabolism of omega-3 and omega-6 essential fatty acids.

Population	Target	Dose	Reference
Healthy adults	1o Prevention of disease (i.e. CVD)	500 mg/d EPA+DHA	Kris-Etherton 2007
Vegetarian & Vegan Diets	1o Prevention of disease (i.e. CVD)	2-4 g/d ALA + 100-300 mg/d DHA	Davis 2003
Pregnancy &	Fetal growth and development	≥ 200 mg/d DHA (can contain EPA)	Koletzko 2007
Lactation	2o CVD Prevention	~ 1 g/d EPA+DHA	Kris-Etherton 2003
Cardiovascular patients	Reduction in blood TAG levels	2-4 g/d EPA+DHA	Miller 2011
Hypertriglyceridemic patients	Depressive related disorders	≥ 1 g/d EPA (can contain DHA)	Martins 2012 / Peet 2005
Neurological health Inflammation (i.e. rheumatoid arthritis)	Pain management	~ 3 g/d EPA+DHA	Kremer 2000 / Calder 2006

Table 1: Dietary recommendations for omega-3 fatty acids in health and disease.

Dietary source	EPA+DHA mg/serving	Amount needed to meet 1000 mg EPA+DHA/day servings/week*	Mean mercury concentration ppm
Finfis			
Tuna – Light, canned	230	30	0.128
Tuna – White (albacore), canned	733	10	0.350
Salmon – Sockeye	673	10	0.022
Salmon – Atlantic, farmed	1825	4	0.022
Salmon – Atlantic, wild	1564	5	0.022
Mackerel – King	341	21	0.730
Mackerel – Pacific	1571	5	0.088
Herring – Pacific	1807	4	0.084
Trout – Rainbow, farmed	744	9	0.071
Trout – Rainbow, wild	840	8	0.071
Halibut – Atlantic & Pacific	200	35	0.241
Cod – Atlantic & Pacific	135	52	0.111
Shellfish			
Shrimp – Mixed species	235	30	0.009
Crab – Alaskan King	351	20	0.065
Lobster – Mixed species	408	17	0.093
Supplements			
Cod liver oil	180	39	ND
Standard fish body oil	300	23	ND
Specialty concentrate	600	12	ND
Pharmaceutical concentrate (Lovaza®)	840	8	ND

Table 2: Amount of EPA and DHA in selected fish/shellfish and fish oil supplements and approximate servings required per week to provide 1000 mg of EPA+DHA per day.

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Questions

1. Which of the following is an essential fatty acid?

- a) alpha linolenic acid (ALA)
- b) eicosapentanoic acid (EPA)
- c) docosahexanoic acid (DHA)
- d) gamma linoleic acid (GLA)

2. Studies show that ALA conversion results in which of the following:

- a) ~5% of ALA is converted to eicosapentaenoic acid (EPA)
- b) 10% of ALA is converted to docosahexaenoic acid (DHA)
- c) under 0.5% of ALA is converted to eicosapentaenoic acid (EPA)
- d) none of the above

3. Essential fatty acids are known to be critical for neurological and cardiovascular health. In particular, ALA and -6 linoleic acid (LA, 18:2 -6) are termed essential fatty acids because humans lack the delta ()15- and 12-desaturase enzymes required for insertion of a double bond at the -3 or -6 position, respectively. Since EPA and DHA can be synthesized from ALA, they are NOT considered essential.

- a) True
- b) False

4. The primary site of ALA desaturation and elongation is the gut, where 90% of conversion occurs.

- a) True
- b) False

5. Which is the rate limiting enzyme in the conversion of ALA to EPA?

- a) 5-desaturase
- b) 6-desaturase
- c) 5-elongase
- d) 6-elongase

6. Which of the following is true about DHA?

- a) it is the predominant -3 PUFA esterified into tissue membrane phospholipids;
- b) it is metabolized to produce resolvins and protectins in the resolution of inflammation;
- c) it is synthesized from EPA via DPA (docosapentanoic acid)
- d) all of the above

7. The US Institute of Medicine's (IOM) and Health Canada have established an Adequate Intake (AI) for ALA as 1.1-1.6 g/day. There is concern however that this level is not sufficient for production of adequate amounts of EPA and DHA, as recommended by professional organizations.

- a) True
- b) False

8. EPA and DHA have a Generally Recognized As Safe (GRAS) status when taken up to 3g per day, including in patients with diabetes, bleeding tendencies, and elevated LDL cholesterol.

- a) True
- b) False

9. Potential problems of relying strictly on dietary sources of EPA and DHA include the following:

- a) Oily fish contain over 1g EPA+DHA per 3oz serving, however white fish may contain only 0.14g;
- b) Two servings per day are required in order to achieve therapeutic serum levels;
- c) High levels of fish consumption may lead to excess exposure to heavy metals such as mercury;
- d) all of the above

10. The following nutrients are cofactors of the 5- and 6-desaturase enzymes: vitamin A, vitamins B3, B6, and C, magnesium and zinc.

- a) True
- b) False

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